THE ABILITY OF LACTOBACILLUS PLANTARUM BSL IN REDUCING THE TISSUE DAMAGE OF LIVER AND SPLEEN IN RATS INFECTED BY LISTERIA MONOCYTOGENES ATCC 7644

Firat Meiyasa^{12*}, Betty Sri Laksmi Jenie³, Lilis Nuraida^{3,4}, Nurbety Tarigan²

1)Study Program of Food Science, IPB University, Bogor 16680
2)Study Program of Aquatic Products Technology, Christian University of Wira Wacana Sumba,
Waingapu 87113
3)Department of Food Science and Technology, IPB University, Bogor 16680
4)Southeast Asian Food and Agricultural Science and Technology (SEAFAST) Center, IPB University,
Bogor 16680

*Corresponding author (fmeiyasa@yahoo.com)

Abstrak

Lactobacillus plantarum BSL merupakan hasil isolasi dari asinan kubis. Tujuan dari penelitian ini adalah untuk mengevaluasi kemampuan *L. plantarum* BSL dalam mereduksi kerusakan jaringan hati dan limpa pada tikus yang diinfeksi oleh *Listeria monocytogenes* ATCC 7644. Kelompok perlakuan tikus menerima 0,5 kultur suspensi (10° CFU/mL) *L. plantarum* BSL dan kelompok kontrol menerima 0,5 mL NaCl 0,85% setiap hari selama sembilan hari percobaan. Kedua kelompok tikus diinfeksi pada hari ke-3 dengan 0,5 mL kultur suspensi (10°CFU/mL) *L. monocytogenes*. Pada hari ke-2 (sebelum diinfeksi), ke-5, ke-7, dan ke-9 (setelah diinfeksi), tikus dibedah. Selanjutnya, hati dan limpa tikus diuji secara histologi. Hasil penelitian menunjukkan bahwa pemberian *L. plantarum* BSL dapat mereduksi kerusakan hati dan limpa tikus selama percobaan.

Kata kunci: Lactobacillus plantarum BSL; Listeria monocytogenes; Probiotik

Abstract

Lactobacillus plantarum BSL, previously isolated from Indonesian sauerkraut. In this study, we investigated the ability of L. plantarum BSL in reducing the tissue damage of liver and spleen in rats infected by Listeria monocytogenes ATCC 7644. Treatment group of rats received 0.5 mL culture suspension (10° CFU/mL) of L. plantarum BSL and control group received 0.5 mL of 0.85% w/v NaCl daily for nine days of experiment. Both groups were infected at 3rd day with 0.5 mL of suspension of L. monocytogenes (10° CFU/mL). At the 2nd (before infection), 5th, 7th, and 9th day (after infection), the rats were sacrificed and then, liver and spleen were assessed for histopathological. Our study revealed that the administration of L. plantarum BSL could be able to reduce the liver and spleen damage of the experimental rats.

Keywords: Lactobacillus plantarum BSL; Listeria monocytogenes; Probiotic

1. INTRODUCTION

Listeria monocytogenes is a foodborne pathogen which may lead to serious clinical diseases with a mortality rate of 20% - 30% (Lee et al. 2014; Jordan & McAuliffe 2018). Listeriosis is a series of diseases caused by the bacteria L. monocytogenes. There are two main types of listeriosis: a non-invasive form and an invasive form. Non invasive listeriosis (febrile listerias gastroenteritis) is a mild form of the disease affecting mainly otherwise healthy people. Symptoms include diarrhea, fever, headache and myalgia. Invasive listeriosis is a

more severe form of the disease and affects certain high risk groups of the population. These include pregnant women, patients undergoing treatment for cancer, AIDS and organ transplants, elderly people and infants (WHO 2018; Tebib *et al.* 2018). Cases of listeriosis result from consumption of contaminated food, particularly ready-to-eat foods, such as dairy products, smoked fish, and seafood (Allen *et al.* 2016).

Probiotics are live microorganisms which when administered in adequate amounts confer

a health benefit to the host (FAO/WHO, 2002a). The positive effects of probiotics are modulation of immune function, protection from diabetes mellitus, prevention of asthma, protection of barrier function in intestinal epithelial cells, anti-allergy and also as an anti-inflammatory (Gobbetti and Minervini 2014; Yu et al. 2017).

Lactobacillus plantarum BSL isolated from sauerkraut has resistance to 5% bile salt and pH 2.5. L. plantarum BSL also has antagonistic properties against pathogen such as B. cereus, S. aureus, and E. coli. Moreover, L. plantarum BSL were able to reduce total cholesterol of rat blood serum, enhance lactobacilli growth, and suppress coliform and staphylococci in rat faeces (Kusumawati et al. 2003; Kusumawati et al. 2008). In addition, Meiyasa et al. (2017) reported that administration of L. plantarum BSL significantly increased the population of and reduced the number of L. monocytogenes compared to control group either in the faeces, caecum, or caecum content, and it effectively reduced the liver and spleen tissues damage in rats. Numerous studies have reported that probiotics have a beneficial effect on the host. However, the role of probiotics in reducing tissue damage of liver and spleen in rats infected by monocytogenes little has been reported.

Based on this, that study aims to evaluate the ability of *L. plantarum* BSL in reducing tissue damage of liver and spleen in rats infected by *L. monocytogenes*.

2. MATERIAL AND METHODS

2.1 Rats

Sprague-Dawley rats (*Rattus norvegicus*) were purchased from Faculty of Animal Science, IPB University. A total of 60 rats (40days old) were randomly allocated and housed in cages (one rat per cage). The rats were housed under standard conditions in the barrier unit: light/dark schedule was constant at 12/12 h and humidity at 50–60% (de Waardet al. 2002). All experiments were approved by Animal Ethical Committee of Bogor Agricultural University.

2.2 Diets

Diet of 20 g/rat/day of feed was formulated according to AOAC (2005) consisted of casein,

corn oil, mineral mix, vitamin mix, carboxymethyl cellulose (CMC), water, and corn starch.

2.3 Bacterial Culture

L. plantarum BSL was obtained from Laboratory of Food Microbiology, Department of Food Science and Technology, IPB University. L. plantarum BSL was grown in MRS broth (Oxoid CM359) at 37 °C for 24 h, followed by pour plating in MRS agar (Oxoid CM361) incubated at 37 °C for 24-48 h for counting the bacterial cells. L. monocytogenes strain ATCC 7644 was cultured BHI broth (Oxoid CM1136) incubated at 37 °C for 24 h, and then counted in Listeria Selective Agar Base (Oxoid CM856), after incubation at 37 °C for 24-48 h (Meiyasa et al. 2018).

To prepare cell suspension, a 24 h culture of *L. plantarum* BSL in MRSB was centrifuged (2000 g, 10 min, 4 °C) and the cell pellet was resuspended in 10 mL volume of 0.85% NaCl. The number of cells of *L. plantarum* BSL in suspension was enumerated in MRS agar. Similar method was applied for *L. monocytogenes* ATCC 7644, only the media was used BHI broth (Meiyasa *et al.* 2018).

2.4 In vivo experiment of antilisterial activity (de Waard et al. 2002)

Rats having initial weight of 100-140 g were acclimatized for 1 week and received a standard diet. In anti-listerial experiment, a total of 48 rats were divided into 2 groups (24 rats for treatment group and 24 rats control group).

L. plantarum BSL and *L. monocytogenes* ATCC 7644 were grown in MRS broth and BHI broth respectively for 24 h at 37 °C. This culture were centrifuged (2000 g, 10 min. 4 °C) and resuspended in 0.85% NaCl in order to obtain cell number up to 10⁹ CFU/mL. This suspension was then administered to rats by the oral route.

The treatment group was administered with *L. plantarum* BSL culture (10⁹ CFU/mL), while the control group was administered with 0.85% NaCl (0.5 mL/rat/day) for 9 days (day-0 – day-9, except at day-3). Rats were then infected by 0.5 mL of *L. monocytogenes* ATCC 7644

The rats were sacrificed by cervical dislocation in 2ndday (before infected by *L. monocytogenes*), 5th day, 7th day, and 9th day (after infected by *L. monocytogenes*) to obtain for histopathology analysis, rat spleen and liver

were stained using Hematoxylin-Eosin (HE) and observed the cell changes under light microscope. Data were evaluated using the descriptive analysis.

3. RESULTS AND DISCUSSION

Histological analysis on liver such as perivascular macrophages (PVM), fatty degeneration (FD), sinusoidal dilatation (SD), and central venous of vibrosis (CVV). Based on the research results show that descriptively administration of *L. plantarum* BSL could reduce tissue damage of liver compared with the control group (Table 1). It is seen that before rats infected by *L. monocytogenes* (in day-2) there was no change in the PVM and FD. However, there has been a change on the SD and CVV in rats liver either in control or treatment group.

Administration of *L. plantarum* BSL for nine days after rats infected by L. monocytogenes showed that could be able to reduce the level of damage in rats liver tissue, especially in the CVV. On the other hand, tissue damage of rats liver in the control group were still observed with the level of minor (+) and moderate (++) damage approximately 67% of the rat population (4/6). In addition, administration of L. plantarum BSL could be able to reduce damage of rats liver such as, PVM and FD approximately 33% of the rat population (2/6) and SD of about 17% of the rat population (1/6) with extent of minor damage (+). In contrast, control group at ninth day were still observed and even more severe as PVM and FD with damage level minor (+), moderate (++) and heavy (+++) approximately 67% of the rat population (4/6). Furthermore, SD with the extent of damage minor (+) and moderate (++) approximately 50% of the rat population (3/6).

Spleen damage in rat was observed including lymphoid glands (LG), increased macrophage cells (IMC), secondary lymphoid cells (SLC) and increased giant cells (IGC) (Table 2). Damage of rat spleen also observed as lymphoid glands, increased macrophage cells, secondary lymphoid cells and increased giant cells were detected at two days, before infected with L. monocytogenes in both the the experiment group. control and Administration of L. plantarum BSL for nine days could be able to reduce spleen tissue damage in rats compared with the control group. For nine days of treatment with *L. plantarum* BSL could be able to reduce tissue damage in rats spleen. However, tissue damage in rats spleen were still observed at nine days.

In rats, administered with *L. plantarum* BSL, five rats (5/6) appear normal again either in the LG, SLC, IMC or IGC with one of six rats suffered minor damage (+). In contrast to the treatment group, in the control group most of the rats were damaged. Seen in the LG and SLC with extent of the damage minor (+) and moderate (++) approximately 67% of the rat population (4/6). Moreover, IMC and IGC also increased by about 50% of the rat population (3/6). Based on this, it can be concluded that the administration of *L. plantarum* BSL for nine days was able to reduce tissue damage of ratsliver and spleen compared with the control group.

In agremment with a previous study reported by Bambirra et al. (2007), that treatment with Lactobacillus sakei 2a could be able to reduce damage lesions and also destruction of the mucosa with infiltrate of inflammatory cells, predominantly macrophages and neutrophil in rats infected by L. monocytogenes Scott A. Furthermore, dos Santos et al. (2011) reported that treatment with L. delbrueckii UFV-H2b20 significantly reduced the damage of liver such as necrosis and degenerative changes of hepatocytes lower than the control group in rats infected by L. monocytogenes 10403S. Other than that, L. plantarum B7 was able to reduce liver translocation and ameliorates histopathological lesions in rats infected by S. thphimurium (Tsai et al. 2005), and then Acurcio et al. (2017) also reported that administration of L. plantarum B7 significantly extent of ileal villi heights and number of hepatic inflammatory foci significantly lower than the control group. In addition, Meiyasa (2017) reported that treatment with *L. plantarum* BSL during nine-days could be able to reduce damage of liver and spleen in rats such as edema, necrosis, cell infiltration of lymphoid, proliferation of bile tract, vacuolization of hepatocytes cell for liver and depletion of lymphoid cells, extension of pulp area, and cell proliferation for spleen.

Administration of *L. plantarum* BSL can reduce tissue damage of liver and spleen in rats possibly through mechanisms such as the production of antagonistic compounds and

inhibitory substances, protective effects against pathogens (Acurcio et al. 2017; Servin 2004; Prieto et al. 2015 competition for nutrients or adhesion sites and/or immunomodulation (Bambirra et al. 2007; Gibson et al. 2005), cytokine production (Castanheira et al. 2007), and reduces pro-inflammatory interleukin secretion (Prieto et al. 2015).

4. CONCLUSION

In summary, our findings demonstrate that *L. plantarum* BSL is able to protect the rats that associated with the ability in reducing the tissue damage of liver and spleen in rats which infected by *L. monocytogenes*.

5. REFERENCES

- Acurcio, L.B., Bastos, R.W., Sandes S.H D.C., Guimaraes, A.C.D.C., Alves, C.G., dos Reis, D.C., Wuyts, S., Nunes, A.C., Cassali, G.D., Lebeer S., de Souza, M.R., Nicoli, J.R. 2017. Protective effects of milk fermented by *Lactobacillus plantarum* B7 from Brazilian artisanal cheese on a *Salmonella enterica*serovar Typhimurium infection in BALB/c mice. *Journal of Functional Foods*. 33, 436 445.
- Allen, K.J., Wałecka-Zacharska, E., Chen, J.C., Kosek-Paszkowska, K., Devlieghere, F., Van Meervenne, E., Osek, J., Wieczorek, K., Bania, J. 2014. *Listeria monocytogenes* Anexamination of food chain factors potentially contributing to antimicrobial resistance. *Food Microbiology*, doi: 10.1016/j.fm.2014.08.006.
- [AOAC] Association of Official Analytical Chemist. 2005. Official Methods of Analysis. Washington, DC.
- Bambirra, F.H.S., Lima, K.G.C., Franco, B.D.G.M., Cara, D.C., Nardi, R.M.D., Barbosa, F.H.F., Nicoli, J.R. 2007. Protective effect of Lactobacillus sakei2a against experimental challenge with Listeria monocytogenes in gnotobiotic mice. Journal Society for Applied Microbiology. 45(6): 663-667.
- Castanheira, L.G., Castro, J.M.A., Martins, F.O.A., Nicoli, J.R., Vieira, L.Q., Afonso,

- L.C.C. 2007. *Lactobacillus delbrueckiias* a potential skin adjuvant for induction of type 1 immune responses. *Frontiers in Bioscience*.12: 1300-1307.
- de Waard, D., Garssen, J., Bokken, G.C.A.M., Vos, J.G. 2002. Antagonistic activity of Lactobacillus casei strain Shirota against gastrointestinal Listeria monocytogenes infection in rats. International Journal of Food Microbiology. 73(1): 93-100.
- dos Santos, L.M., Santos, M.M., Humberto, P.S.S., Rosa, M.E.A., Jacques, R.N., Leda, Q.V. 2011. Monoassociation with probiotic Lactobacillus delbrueckii UFV-H2b20 stimulates the immune system and protects germfree mice against Listeria monocytogenes infection. Medical Microbiology and Immunology. 200(1): 29-38.
- Food and Agricultural Organization (FAO)/World Health Organization (WHO). 2002a. Health and nutritional properties of probiotics in food including powder milk with live lactic acid bacteria e Joint Food and Agricultural Organization of the United Nations and World Health Organization Expert Consultation Report. Córdoba, Argentina.
- Gibson, G.R., McCartney, A.L., Rastall, R.A. 2005. Prebiotics and resistence to gastrointestinal infections. *British Journal of Nutrition*. 93: 31-34.
- Gobbetti, M and Minervini, F. 2014. *Lactobacillus casei*. Encyclopedia of Food Microbiology. 2: 432-438.
- Jordan, K and McAuliffe, O. 2018. Listeria monocytogenes in foods. Advances in food and nutrition research. 86:181-213.
- Kusumawati, N., Jenie, B.S.L., Setyahadi, S., Hariyadi, R.D. 2003. Selection indigenous lactic acid bacteria as probiotic strain with the ability to lower cholesterol. *Journal of Microbiology Indonesia*. 8(2): 39-43.
- Kusumawati, N., Jenie, B.S.L., Setyahadi, S., Hariyadi, R.D. 2008. Antibacterial activity of Lactobacilli fermented food origin Indonesia against pathogens and their effects on the

- intestinal microflora rats. *Journal of Medicine of Natural Materials*. 7(1): 69-75.
- Lee, D.K., Park, J.E., Kim, M.J., Seo, J.G., Lee, J.H., Ha, N.J. 2014. Probiotic bacteria, *B. longum*and *L. acidophilus*inhibit infection by rotavirus in vitro and decrease the duration of diarrhea in pediatric patients. *Clinics and Research in Hepatology and Gastroenterology Journal*. 39(2): 237-244.
- Meiyasa, F. 2017. Evaluasi Efektivitas Lactobacillus plantarum BSL dalam Menghambat Infeksi Listeria monocytogenes secara in vivo (Magister Thesis, Bogor Agricultural University (IPB).
- Meiyasa, F., Jenie, B.S.L., Nuraida, L., Wahyuwardani, S. 2018. Effectivity of Lactobacillus plantarum BSL against Listeria monocytogenes in rats. Malaysian Journal of Microbiology. 14(3): 282-292.
- Prieto, S.D.M., Moreno, E., Meca, T.B., Lasserrot, A., Lopez, M.D.R., Bravo, A.R., Valera, M.J. 2015. A. Lactobacillus plantarum strain isolated from kefir protect against intestinal infection with Yersinia enterocolitica O9 and modulates immunity in mice. Research in Microbiology. 166(8): 626-632.
- Puertollano, E., Puertollano, M.A., Chamorro, LC., de Cienfuegos, G.A., Bravo, A.R., de Pablo, M.A. 2008. Orally administered *Lactobacillus plantarum* reduces pro-

- inflammatory interleukin secretion in sera from *Listeria monocytogenes* infected mice. British Journal of Nutrition. 99(4): 819-825.
- Servin, A.L. 2004. Antagonistic activities of Lactobacilli and Bifidobacteria against microbial pathogens. FEMS Microbiology Reviews. 28(4): 405-440.
- Tebib, N., Bill, O., Niederhauser, J., Christin, L. 2018. Case study: An uncommon complication of *Listeria monocytogenes* infection: Polyradiculoneuritis following Listeria meningoencephalitis. *IDCases*. 12: 101-103.
- Tsai, C.C., Hsih, H.Y., Chiu, H.H., Lai, Y.Y., Liu, J.H., Yu, B., Tsen, H.Y. 2005. Antagonistic activity against *Salmonella* infection in vitro and in vivo for two *Lactobacillus* strains from swine and poultry. *International Journal of Food Microbiology*. 102(2): 185-194.
- [WHO]. 2018. Media Centre. Listeriosis, Fact Sheet.http://www.who.int/mediacentre/facts heets/listeriosis/en/
- Yu, J., Zhu, S.H., Yang, G.Y., Zhang, W., Zhou, D., Su, J.H., Wang, J.F. 2017. Anti-inflammatory capacity of *Lactobacillus rhamnosus* GG in monophasic variant Salmonella infected piglets is correlated with impeding NLRP6-mediated host inflammatory responses. *Veterinary Microbiology*. 210: 91-100.

Table 1. Reduction of liver damage during nine-day administration of *L. plantarum* BSL.

Group	Numb er of rats		Days														
		2			5				7				9				
			Liver														
		PVM	FD	SD	CVV	PVM	FD	SD	CVV	PVM	FD	SD	CVV	PVM	FD	SD	CVV
Control	1	-	-	-	-	+	++	-	+	-	-	+	+	-	-	-	+
	2	-	-	-	-	-	+	-	+	+	++	++	-	++	+++	++	-
	3	-	-	-	-	+++	-	-	-	++	++	+	++	+++	++	++	++
	4	-	-	-	-	+++	++	++	-	+	+	-	-	+	-	-	+
	5	-	-	-	+	+	-	+	++	+++	+	+	+	+	++	+	-
	6	=.	+	-	+++	+	+++	++	++	+	+++	-	+	-	++	ı	+
Treatment	1	=.	-	-	-	-	-	++	+	++	-	+	-	-	-	ı	
	2	-	-	-	-	+++	+++	+	-	++	++	-	-	-	-	-	-
	3	-	+	-	-	++	-	-	+	-	++	-	+	+	-	-	-
	4	-	++	-	-	+++	++	+	++	+	+	-	-	+	+	-	-
	5	-	-	-	-	+	+	-	-	-	-	+	+	-	-	-	-
	6	-	-	-	-	+	-	+	+	++	-	-	-	-	+	+	-

Note: Damage level of liver tissue: no (-); minor (+); moderate (++); heavy (+++)

Table 2. Reduction of spleen damage during nine-day administration of L. plantarum BSL

Group	Numb er of rats		Days														
		2				5			7				9				
			Spleen														
		LG	IMC	SLC	IGC	LG	IMC	SLC	IGC	LG	IMC	SLC	IGC	LG	IMC	SLC	IGC
Control	1	-	-	-	-	++	++	-	-	-	+	+	++	+	++	-	-
	2	-	-	-	-	-	-	-	-	++	++	++	+++	+	-	++	++
	3	+	-	-	-	+	++	-	+	++	++	+	-	-	++	+	+
	4	-	-	-	-	-	-	++	+++	-	-	-	-	++	-	+	-
	5	-	-	-	-	++	+	+	+	+	+++	+	+	+	-	++	++
	6	-	+	-	-	•	-	++	+	-	-	-	+	-	++	-	-
	1	-	+	-	+	+	-	-	-	-	-	-	-	-	-	+	-
Treatment	2	-	-	-	-	-	+	-	++	+	-	-	-	-	-	-	-
	3	-	+	-	-	+	-	++	-	-	+	+	+	-	-	-	-
	4	-	-	-	-	-	++	++	++	-	-	+	-	-	-	-	-
	5	-	-	-	-	-	-	+++	-	-	+	++	+	-	-	-	+
	6	-	++	-	-	++	+	-	+	-	-	-	+	+	+	ı	-

Note: Damage level of spleen tissue: no (-); minor (+); moderate (++); heavy (+++)